

## Potential of Purple Sweet Potato (*Ipomoea batatas L*) To Increase BDNF Level and VEGF Expression in The Cerebellum of Ischemic Stroke Rats

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Received 18 August 2017; Revised 4 January 2018; Accepted 5 January 2018

### ABSTRACT

This paper reports the effect of anthocyanin from purple sweet potato in the cerebellum of ischemic stroke rats model by Middle Cerebral Artery Occlusion (MCAO) technique. This technique was carried out by ligating the blood flow in External Carotid Artery (ECA), Common Carotid Artery (CCA) and Internal Carotid Artery (ICA) for 3 hours, followed by reperfusion. The MCAO technique proved a technique for preparing ischemic stroke rats. This technique induces releasing of Brain-Derived Nuclear Factor (BDNF) and Vascular Endothelial Growth Factor (VEGF) in the cerebellum of ischemic stroke rats. The levels of BDNF measured by ELISA technique and VEGF expression used immunohistochemistry technique. The results showed that anthocyanin from purple sweet potato increases the level of both BDNF and VEGF expression in the cerebellum of ischemic stroke rats. It is suggested that anthocyanin could be used as a therapeutic agent for ischemic stroke.

Key word: purple sweet potato, BDNF, VEGF, MCAO, ischemic stroke, cerebellum

### INTRODUCTION

Anthocyanin, a class of flavonoid, is distributed in plant tissues such as leaves, roots, and seeds. This compound is correlated to red, blue and purple color in plants[1]. Among other plants like strawberry, perilla, and red cabbage, purple sweet potato contains the highest anthocyanin levels[2]. In addition, due to its natural food pigment, high nutrition, and health benefits, this compound has been suggested as a source of healthy food recommendation for preventing diseases[3]. Anthocyanin from purple sweet potato has various biological activities, including as an antioxidant, anti-inflammation, anticancer, antimutagenic, and anti-hyperglycemic[3,4]. Biological activities of anthocyanin attenuate insulin resistance levels[5], suppress the liver fibrosis[6], against hepatotoxicity induced *tris*-BHP[7], and prevent acute and sub-acute alcoholic liver damage[8]. Recent studies have shown that anthocyanin from purple sweet potato inhibits inflammation response in rat brain induced lipopolysaccharide by blocking of signaling pathways in Extracellular Signal-Regulated Kinase (ERK), The C-Jun N-Terminal Kinase (JNK) and Nuclear Factor Kappa-B (Nf-kB) signaling[9]. As flavonoids derivative, anthocyanin has been predicted to maintain the fragility of capillaries to protect the brain from injury[10]. Thus, due to its biological activities, anthocyanin from purple sweet potato could be used as a therapeutic agent in brain ischemic stroke.

The journal homepage [www.jpacr.ub.ac.id](http://www.jpacr.ub.ac.id)  
p-ISSN : 2302 – 4690 | e-ISSN : 2541 – 0733

Ischemic stroke is caused by partial or complete occlusion of the cerebral artery which activating ischemic cascade[11]. This process induces cytotoxicity, activation of inflammation response, oxidative damage and ionic imbalance. These processes induce neuron damaging. In contrast, the hypoxia caused by blood flow blocking triggers a protein stress to form Heat Shock Protein-90 (HSP-90)[12]. This protein stimulates BDNF activation through activation of protein kinase B and Adenine Monophosphate-Activated Protein Kinase (AMPK)/ERK[13]. The formation of Brain-Derived Nuclear Factor (BDNF) is started with modification of phosphorylation protein component at synapses which modify a translation activity in the transcription process. It is also a protective sign of the brain that sprouting, growth, survival, and differentiation of neuronal structure and synapses after ischemic stroke [14]. Besides that, hypoxia stimulates producing Hypoxia-Inducible Factor-1 alpha (HIF-1 $\alpha$ ); a protein that responsible for VEGF expression. VEGF itself is a mediator of angiogenesis activated Phosphoinositide 3-Kinase (PI3K)/Akt and Mitogen-Activated Protein Kinase (MAPK) signaling pathways. Both pathways play a role in survival, proliferation, and migration of endothelial cells microvascular[15]. VEGF which bind to a receptor in endothelial cell surface stimulate angiogenesis responses, in addition, to activate, proliferate, migrate, and survive of neurons [16]. However, the increase of VEGF expression itself is the key to a protective effect in neuron damaging.

This research is focused on the cerebellum. This region does not affect directly with ischemic stroke rats induced by MCAO but it is known that the function of the human body depends on the interaction of all organs, and brain damage resulting from a stroke can lead to multiple organ failures especially in cerebellum brain area[12]. The correlation between MCAO and cerebellum was not fully understood. Thus, this paper aimed to report the differences in BDNF levels and VEGF expression in the cerebellum of ischemic stroke rats.

## EXPERIMENT

### Chemicals and instrumentation

The sample of purple sweet potato used in this experiment were brought from a commercial product (Sweet IMO) harvested from Bali. The chemicals used are ketamine, xylazine, PBS pH 7, PBS-Tween : PMSF, Tris-HCl, hydrogen peroxide 1 %, primer antibody of anti VEGF, secondary antibody anti-rat labeled streptavidin-biotin, SA-HRP, Meyer hematoxylin Eosin, water and Elisa Kit Rat BDNF from Elabscience Biotechnology Co., Ltd (Wuhan, Hubei Province, China) Catalog No: E-EL-R1235.

Instrumentation used for analysis the purple sweet potato beverage was UHPLC ACCELL type 1250 (Thermo Scientific). This instrumentation is equipped with an electrospray ionization (ESI) with the positive mode. A sample of purple sweet potato was injected into column equilibrated in 95:5 A: B. A= 0.1 % formic acid in water; B= 0.1 % formic acid in acetonitrile and flow rate = 300  $\mu$ l/min.

### LC-MS analysis

The anthocyanins in purple sweet potatoes (*Ipomoea batatas*) was separated and identified the structure by LC-MS/MS.

### Preparation of experimental animals

An adult male *Rattus norvegicus* aged 3 months with the weight of  $300 \pm 30$  g. Animals are kept at the clean standard of a laboratory animal for 7 d to adapt to the environment in the Animal Experimental Laboratory of Biosains Institute. They were maintained at a temperature of 25 °C and relative humidity with a 12-h light/12-h dark cycle and they were

provided with free access to food and water. All condition and handling conducted based on ethical protocols approved by Brawijaya University Animal Ethics Committee No. 651-KEP-UB. In this experiment, all animals were divided into 5 groups. The first was a control group (KN). The second was a group of 1 h ischemic stroke (K1). The third was a group of 72 h ischemic stroke. The fourth was a therapy group of 24 h ischemic stroke (T1) and the last was a therapy group of 72 h ischemic stroke. The therapy used purple sweet potatoes was given 2 mL by oral each day.

### **Middle cerebral artery occlusion (MCAO) technique**

The ligation of the middle cerebral artery was conducted based on the conventional method with slight modification[17]. This method was done by ligating the brain artery in the Common Carotid Artery (CCA), External Carotid Artery (ECA) and Internal Carotid Artery (ICA). The occlusion was done for 3 h. After 3 h, the ligation was released for reperfusion. As a result of ischemic reperfusion, the ischemic stroke will occur caused by lack of oxygen and glucose to the brain.

### **Isolations of cerebellum protein**

The cerebellum was mixed and added with PBS-Tween: PMSF. The mixture was homogenized, sonicated and centrifuged. The homogenate was added with ethanol absolute and left overnight. The supernatant was centrifugated for 15 min at 10.000 rpm. The sample was taken and dried. The precipitate was mixed with Tris-HCl pH 6,8 with the ratio 1:1 (w/v)[18].

### **BDNF levels**

The levels of BDNF were determined using Rat BDNF Elisa Kit (Elabscience Biotechnology Co, Ltd, Guangdong Science, and Technology, China) based on manufacturer instructions. All the cerebellum of rat tissues was prepared for the analysis. The cerebellum was isolated and prepared for Elisa Kit test. 100  $\mu$ L of standard, blank and sample were added per-well, then were incubated for 90 minutes at 37  $^{\circ}$ C. The liquid has removed the liquid and 100  $\mu$ L of biotinylated detection Ab was added and was incubated for 1 hour at 37  $^{\circ}$ C. The mixture was aspirated and washed three times with 100  $\mu$ L wash buffer. Next, 100  $\mu$ L of HRP conjugate was added to the mixture, and this was incubated for 30 minutes at 37  $^{\circ}$ C. This solution was aspirated and washed again wash buffer. A 90  $\mu$ L of substrate solution was added to the solution and incubated for 15 minutes at 37  $^{\circ}$ C. In order to stop the reaction, 50  $\mu$ L stop solution was added. The optical density of the sample solution was determined using microplate reader at a wavelength of 450 nm. The amount of BDNF was calculated from the standard calibration curve.

### **Immunohistochemistry of VEGF**

Rats were sacrificed and cerebellums were quickly removed. Cerebellum sections were deparaffinized using xylene, rehydrated through a graded series of ethanol. The sections were placed in an oven for 7 days for immunohistochemistry processing. The slides were treated with PBS for 15 min and treated with 1% hydrogen peroxide for 20 min. Then, the slides were blocked using BSA 1%. The slides were incubated with primary antibodies anti-VEGF overnight at room temperature. After a thorough rinse in PBS, the biotin-labeled secondary antibody was applied for 1 h. The sections were treated with SA-HRP for 1 h at room temperature and visualized by treating DAB for 40 min then counterstained with Meyer's

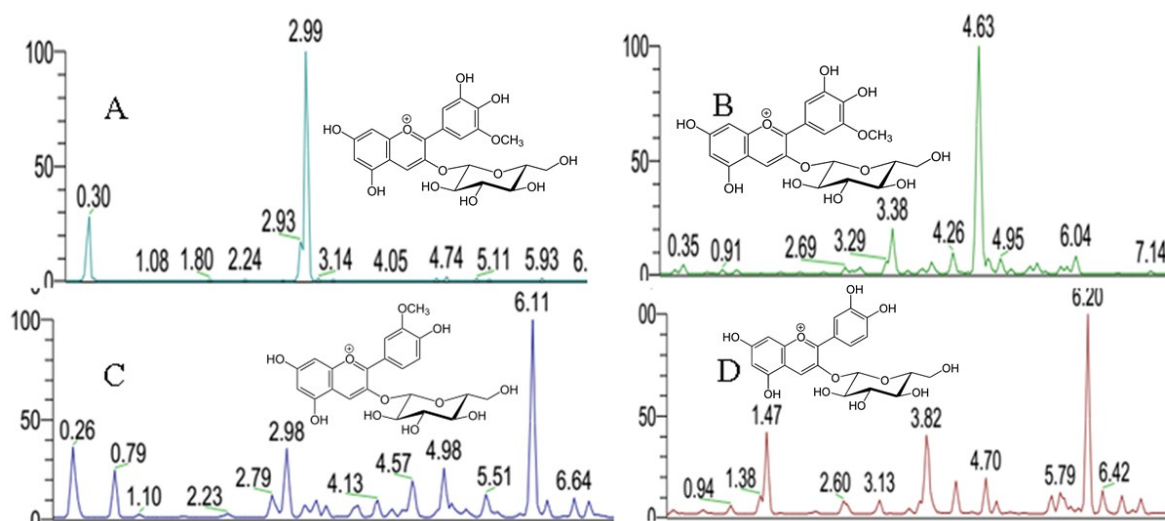
hematoxylin-eosin. The slides were covered and observed using a microscope with magnificent 400X.

## RESULT AND DISCUSSION

**Figure 1** shows the LC-MS chromatogram of purple sweet potato beverages. The data shows purple sweet potato beverages contain high anthocyanin. The anthocyanins predicted are petunidin, cyanidin, and malvidin derivatives. Anthocyanins are known to be an antioxidant as the free radical scavenger and give an electron to stabilize the activity of free radicals [19]. Thus, the purple sweet potato beverages can use as therapeutic in stroke ischemic treatment.

**Table 1.** Prediction of anthocyanins structure in purple sweet potatoes beverages using LCMS

The prediction of anthocyanine content	Retention time (min)	[M] <sup>+</sup> (m/z)	Fragment ion (m/z)
Petunidin-3,5-O-diglucoside	2.99	625	316.5-317.5
Peonidin-3-O-glucoside	4.63	463	300.5-301.5
Delphinidin-3-O-glucoside	6.11	465	302.5-303.5
Cyanidin-3-O-glucoside	6.20	595	286.5-287.5



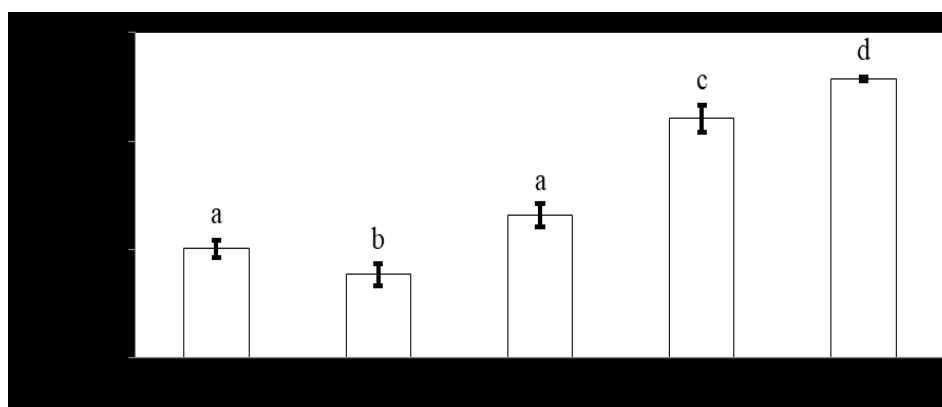
**Figure 1.** LCMS chromatogram of anthocyanins in purple sweet potato beverages. **A** show of Petunidin-3,5-O-glucoside **B** shows peonidin-3-O-glucoside. **C** shows delphinidin 3-O-glucoside. **D** shows cyanidin-3-O-glucoside.

### The BDNF levels after anthocyanin therapy on Ischemic stroke

In the current work, the potential of anthocyanin in purple sweet potato as a brain neuroprotection with ischemic stroke disorder was discussed. The discussion based on changes in BDNF cerebellum levels. In control group, BDNF concentrations were  $420.05 \pm 31.63$  pg/mg. However, at 1 h after ischemic reperfusion, there was a decrease in BDNF levels to 23.80%. BDNF levels showed a change after 72 hours of reperfusion ischemia. At 72 hour of after ischemic reperfusion, BDNF increased to 30.98%. Increasing of BDNF production after ischemic stroke suggests neuroprotection and neuronal repairment

naturally[20]. Anthocyanin therapy significantly increases ( $p<0.05$ ) BDNF levels in the cerebellum in ischemic stroke rats. This data suggests that reperfusion ischemic induction resulting in ischemic stroke.

There were significant differences between control group and therapy group in the cerebellum rats with an ischemic stroke. Therapy group shows a significant effect on BDNF concentrations in the cerebellum. The therapeutic group at 24 hours and 72 hours showed elevated BDNF levels to 187.84 % and 326.05 %, respectively. These changes indicate a neuroprotective ability induced by the antioxidant activity that anthocyanin possesses from purple sweet potato. Figure 2 shows a change in BDNF levels in the cerebellum of ischemic stroke rats.



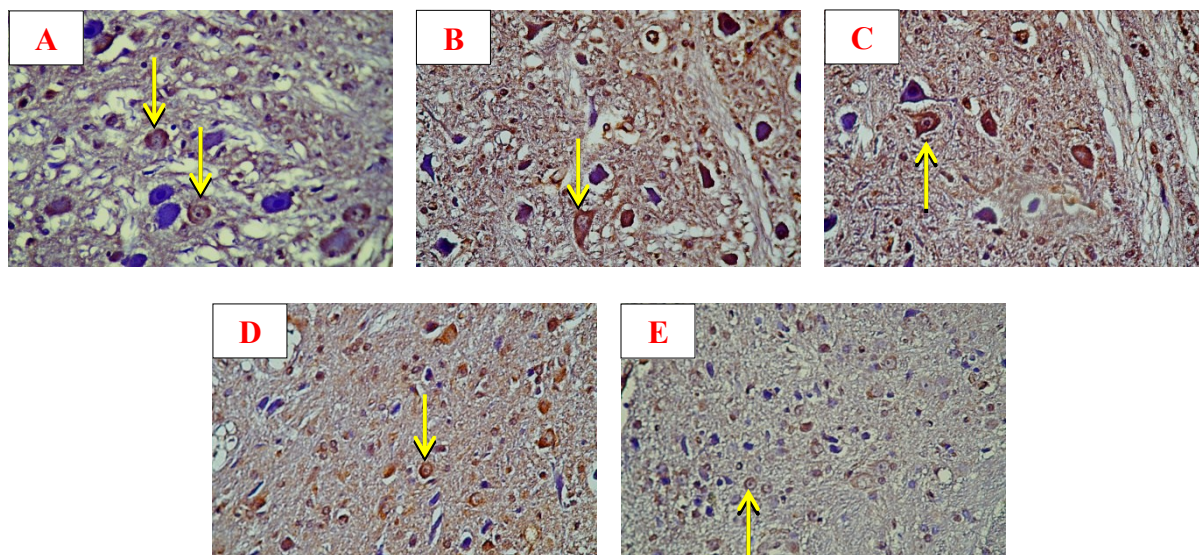
**Figure 2.** Graph of BDNF levels in the cerebellum in an ischemic stroke rats.

The BDNF levels in areas with undifferentiated caused by reperfusion ischemia will increase BDNF expression. This is because BDNF can be produced by both neurons and non-neuron cells. In vitro studies demonstrate the ability of microglial cells, cerebral endothelial cells and astrocytes to express and secrete BDNF when the brain is exposed to ischemic conditions[21]. BDNF may improve neurogenesis after ischemic injury by activating neural and pro-neurogenic pathways PI3K/Akt and MAPK[13].

The anthocyanins contained in the purple sweet potato have the potential as neuroprotective agents to be used to treat ischemic stroke. Neuroprotective action acts in the increase of phosphatidylcholine synthesis, stimulation of glutathione synthesis, and reduce arachidonic acid levels formed after ischemic stroke[10]. Anthocyanin is expected to be able to decrease the cytotoxicity, calcium dysregulation, mitochondrial dysfunction, oxidative and nitrosative stress and inflammation. In addition, anthocyanin acts to reduce tissue acidosis, blood-brain barrier, brain neuron apoptosis, necrosis and autophagy[10].

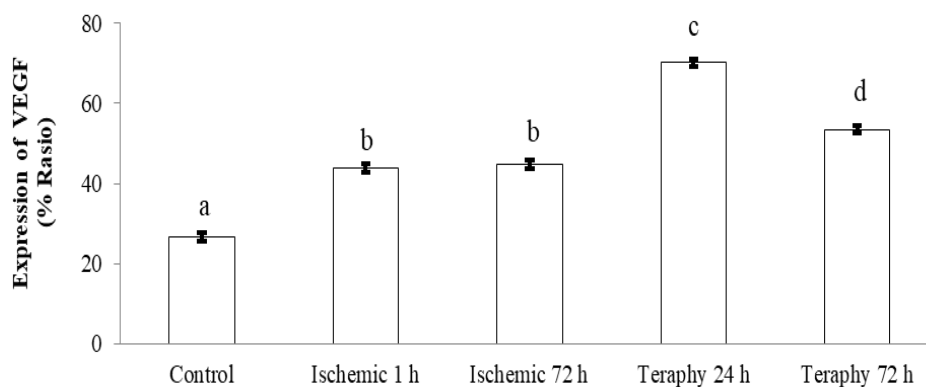
### **Effect of Purple Sweet Potato Anthocyanin on VEGF Expression Of Cerebellum On Ischemic Stroke Rats**

Anthocyanin activity in purple sweet potato is expected to stimulate neurogenesis of nerve cells observed by increasing VEGF expression. Increased VEGF expression is triggered by a reperfusion ischemic stressor by activating PI3K/Akt and MAPK pathways that will enhance the defense and proliferation of endothelial cells. VEGF inhibits ischemic induction through activation of the PI3K signal pathway by increasing the potassium current in the neuron membrane[22]. VEGF expression is observed by the formation of brown color on the cytoplasm of neurons. Figure 3 shows VEGF expression in the control and treatment groups.



**Figure 3.** VEGF expression in rats cerebellum of ischemic rats (magnification 400x)  
A is a control. B group of 1 h ischemic stroke. C group of 72 h ischemic stroke. D therapy group of 24 h ischemic stroke. E therapy group of 72 h ischemic stroke.

The measurement of the percentage of VEGF expression was conducted by the immunoratio program. The program used is ImageJ. It calculates the percentage of DAB-sustained nuclear area (labeling index) and hematoxylin-stained nuclei regions[23]. Based on that measurement, Figure 4 shows VEGF expression on control was  $26.6\% \pm 1.6$ . This value differed significantly in all the groups tested in both K1, K2, T1, and T2. VEGF expression showed a significant increase in groups of rats with ischemic stroke rats group at 1 hour and 72 hours post-ischemic reperfusion with VEGF expression change of  $43.9\% \pm 2.6$  and  $44.8\% \pm 2.7$ . Previous research suggested that VEGF expression may increase within 24 hours after MCAO in neonatal model rats with hypoxic-ischemic injury[24]. A study using albumin as an induced MCAO induced ischemic stroke therapy showed a low VEGF expression at 6 hours and 24 hours post-ischemia. Meanwhile, VEGF expression increased at 24 hours, 72 hours and 60 hours post MCAO[25]. In the study, the highest amount of VEGF expression was demonstrated by the group of treated rats at 24 h post-ischemic reperfusion with a percentage of  $70.2\% \pm 2.6$ . However, VEGF expression decreased to  $53.5\% \pm 3.6$  in the therapy group at 72 hours post-ischemic reperfusion.



**Figure 4.** Graph of VEGF expression in the cerebellum of ischemic stroke rats.

Based on the data, it is known that reperfusion ischemia affects the physiological system of the cerebellum as evidenced by the increase of VEGF expression. In general, the repair and recovery of brain function following the stroke depends on the plasticity of the brain in the non-ischemic part.

## CONCLUSION

This paper shows ischemic stroke affects the cerebellum which shown by changes in BDNF levels and VEGF expression. Increased levels of BDNF and VEGF expression prove that anthocyanin from purple sweet potato has the neuroprotective effect.

## ACKNOWLEDGMENT

This research was supported by dr. Oka Adnyana from Udayana University and Wibi Riawan from Medical Faculty of Brawijaya University. We are thankful for their financial supporting and assistance.

## REFERENCES

- [1] X. He, X. Li, Y. Lv, and Q. He, *Food Sci. Technol*, **2013**, 35(3).
- [2] I. Suda, T. Oki, M. Masuda, and M. Kobayashi, *JARQ*, **2002**, 37(3), 167–173.
- [3] J. Zhao, Q. Yan, L. Lu, and Y. Zhang, *Nutr Res Pract.*, **2013**, 7(5), 359–365.
- [4] Y. Pil, J. Ho, E. Hee, H. Gyun, and J. Wee, *Nutr. Res.*, 2011, 31(12), 896–906.
- [5] Z. Zhang, J. Lu, Y. Zheng, D. Wu, B. Hu, Q. Shan, W. Cheng, M. Li, and Y. Sun, *J. Nutr. Biochem.*, **2013**, 24(6), 1008–1018.
- [7] Y. Pil, J. Ho, J. Min, Y. Chul, and H. Gwang, *Food Chem. Toxicol.*, **2011**, 49(9), 2081–2089.
- [8] H. Sun, T. Mu, X. Liu, M. Zhang, and J. Chen, *J. Agric. Food Chem.*, **2014**, 62(11), 2364–2373.
- [9] Y. Wang, Y. Zheng, J. Lu, G. Chen, X. Wang, J. Feng, J. Ruan, X. Sun, C. Li, and Q. Sun, *Neurochem. Int.*, **2010**, 56(3), 424–430.
- [10] X. Chen and K. Wang, *Acta Pharm. Sin. B*, **2016**, 6(6): 522–530.
- [11] Charleston and R. J. Adam, *Ischemic Stroke Therapeutics: A Comprehensive Guide*, **2016**, Springer, Switzerland.
- [12] S. Ma, H. Zhao, X. Ji, and Y. Luo, *Exp. Neurol.*, **2015**, 272, 41-49.
- [13] C. An, Y. Shi, P. Li, X. Hu, Y. Gan, R. A. Stetler, R. K. Leak, Y. Gao, B. Sun, P. Zheng, and J. Chen, *Prog. Neurobiol.*, **2014**, 115, 6–24.
- [14] R. A. Wardle and M. Poo, *J Neurosci*, **2003**, 23(25), 8722–8732.
- [15] S. Bürger, Y. Yafai, M. Bijl, P. Wiedemann, and R. Schliebs, *Int J Dev Neurosci*, **2010**, 28(7), 597–604.
- [16] K. L. Jin, X. O. Mao, and D. A. Greenberg, *Proc Nati Acad Sci S A.*, **2000**, 97(18), 10242-102427.
- [17] Adnyana IMO, Sudewi AAR, Sumatra DPGP, Suprpta ND, Aulanni'am A., *Bali Med J*, **2017**, 6(1), 156–160.
- [18] R. Setin and C. Mahdi, *J. Pure App. Chem. Res.*, **2013**, 2, 55–61.
- [19] L. P. Gina and C. Mahdi, *J. Pure App. Chem. Res.*, **2016**, 5, 40–47.
- [20] A. Chan, J. Yan, P. Csurhes, J. Greer, and P. Mccombe, *J. Neuroimmunol*, **2015**, 286, 42–47.
- [21] N. Bertrand, P. Garnier, and C. Marie, *Neurochem. Int.*, **2011**, 58 (1), 102–111.
- [22] K. W. Wu, P. Yang, S. S. Li, C. W. Liu, and F. Y. Sun, *Neuroscience*, **2015**, 298, 94–101.

- [23] V. Della Mea, G. L. Baroni, D. Pilutti, and C. Di Loreto, *PLoS ONE*, **2017**, 12 (7), 1–9.
- [24] David A. Greenberg, *Cell Mol Life Sci.*, **2014**, 70 (10), 1753–1761.
- [25] X. Yao, W. Miao, M. Li, M. Wang, J. Ma, Y. Wang, L. Miao, and H. Feng, *Neurosci. Lett.*, **2011**, 472 (3), 179–183.